

Introduction

This protocol shows optimum transfection condition using HilyMax in NIH3T3 cells. To transfect NIH3T3 cells in 24-well plate, follow "Optimum Condition for Transfection" and "Transfection Procedure". When using the other vessel, refer to Table 2 and adjust the amounts of cells, medium, DNA and HilyMax in proportion to the relative surface area.

※Important Note※

Optimum Transfection condition is possibly changed by passage number and culture condition. If transfection efficiency is low by followed this protocol, refer to "Transfected Result by HilyMax" and "Troubleshooting".

Optimum Condition for Transfection (for 24-well plate)

Table 1 Optimum condition for transfection to NIH3T3 cells

| | | |
|----------------------------------|-------------------|-----------------|
| Cell Density | 80% | |
| DNA-HilyMax complex formation | Serum-free medium | 30 μ l |
| | DNA | 0.5 μ g |
| | HilyMax | 1.5-2.5 μ l |
| | Incubation time | 15 min |
| Medium change after transfection | Necessary | |

Transfection Procedure (for 24-well plate)

Cell preparation

- Adjust the concentration of cells to be 80% confluent in 0.5 ml of growth medium prior to transfection.
- Inoculate the cell suspension onto the 24-well plate.
- Incubate cells in CO₂ incubator for 24 hr.

Transfection

- Form the DNA-HilyMax complex
 - Add the serum-free medium(without antibiotics) 30 μ l/well in a sterile plastic tube
 - Add plasmid DNA 0.5 μ g/well and mix by gentle pipetting
 - Add HilyMax 1.5-2.5 μ l/well and mix by gentle pipetting
 - Incubate the mixture of DNA and HilyMax solution at room temperature for 15 minutes
- Add DNA-HilyMax complex to cells in each well and mix by gentle shaking the plate
- Incubate cells in CO₂ incubator for 18-48 hr
- (!) Change the growth medium 4 hours after transfection.

Assay

- Measure protein expression

Transfection in Various Vessels

Table 2 Transfection condition in various vessels

| Culture Vessel | Culture of Cells | | Formation of DNA-HilyMax complex | | |
|----------------|----------------------|----------------|----------------------------------|--------------|-------------------|
| | Surface Area | Plating Medium | Serum-free Medium | DNA | HilyMax |
| 96 -well | 0.3 cm ² | 0.1 ml | 10 μ l | 0.1 μ g | 0.3-0.5 μ l |
| 24 -well | 1.9 cm ² | 0.5 ml | 30 μ l | 0.5 μ g | 1.5-2.5 μ l |
| 12 -well | 3.8 cm ² | 1.0 ml | 60 μ l | 1.0 μ g | 3.0-5.0 μ l |
| 6 -well | 9.2 cm ² | 2.0 ml | 120 μ l | 2.0 μ g | 6.0-10.0 μ l |
| 35 -mm | 8.0 cm ² | 2.0 ml | 120 μ l | 2.0 μ g | 6.0-10.0 μ l |
| 60 -mm | 21.0 cm ² | 5.0 ml | 300 μ l | 5.0 μ g | 15.0-25.0 μ l |
| 100 -mm | 58.0 cm ² | 15.0 ml | 900 μ l | 15.0 μ g | 45.0-75.0 μ l |

Transfected result by HilyMax

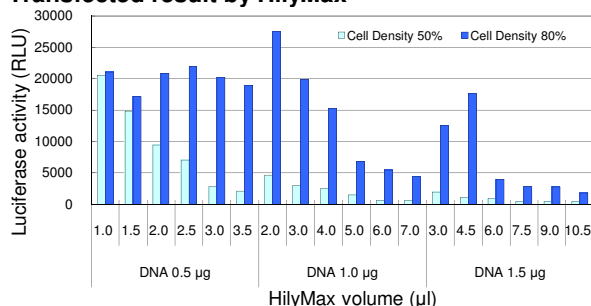


Fig. 1 Transfection Efficiency in NIH3T3 cells

NIH3T3 cells were incubated for 24 hr and transfected pGL3 control vector (Promega) using HilyMax in each conditions. Transfection efficiency (Luciferase activity) was measured in 24 hr after transfection. NIH3T3 cells were cultured in D-MEM medium(Gibco) containing 10%FBS(Gibco) for about 2 weeks after thawing.

50% confluent : 0.9×10^5 cells/well 80% confluent : 1.2×10^5 cells/well

Troubleshooting

-Low Transfection Efficiency-

- Change the DNA(μ g):HilyMax(μ l) ratio to 1:6-1:8.
- Increase the mass of DNA up to 2.0 times and change the DNA(μ g):HilyMax(μ l) ratio to 1:3-1:5.

-High cellular Toxicity-

- Decrease the mass of DNA down to half and change the DNA (μ g):HilyMax(μ l) ratio to 1:3-1:7.

-Check the Material and Condition-

- Was HilyMax Reagent dissolved completely when HilyMax was Prepared?
- Was incubation time of cells after transfection optimum for cells and plasmid?
- Was DNA-HilyMax complex formed in medium without serum and antibiotics?